

CHEMICAL STUDIES ON MEXICAN PLANTS USED IN TRADITIONAL
MEDICINE, III: NEW 4-PHENYLCOUMARINS FROM
EXOSTEMA CARIBAEUM^{1,2}

RACHEL MATA,* FERNANDO CALZADA, MA ROSARIO GARCIA, and MA TERESA REGUERO

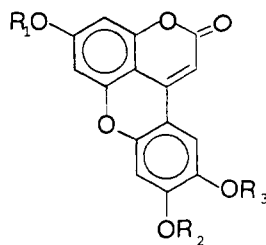
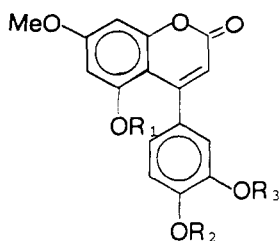
Departamento de Farmacia, División de Bioquímica y Farmacia, Facultad de Química de la Universidad Nacional Autónoma de México, Ciudad Universitaria, Coyoacán 04510, México DF

ABSTRACT.—Investigation of the MeOH extract of *Exostema caribaeum* (Rubiaceae) led to the isolation of three new 4-phenylcoumarins. Their structures, 5-O- β -D-galactosyl-7-methoxy-3',4'-dihydroxy-4-phenylcoumarin [**1a**] 7,4',5'-trihydroxy-4-phenyl-5,2'-oxido-coumarin [**2a**] and 7,4'-dimethoxy-5'-hydroxy-4-phenyl-5,2'-oxido-coumarin [**3a**] were elucidated by spectral methods and chemical transformations. It was also demonstrated that 4-phenylcoumarins undergo oxidative cyclization under basic conditions in the presence of air to give 4-phenyl-5,2'-oxido-coumarins.

Exostema caribaeum (Jacq.) Roem. et Schult. (Syn. *Cinchona caribaea*, Rubiaceae) is a tropical species found in the West Indies, México, and Costa Rica. The stem bark of this plant is used in folk medicine as a substitute for quinine (1-3). Previous phytochemical investigations resulted in the isolation of exostemin and mannitol (4,5). The present work describes the isolation and structure elucidation of three new 4-phenylcoumarins.

Dried stem bark of *E. caribaeum* was extracted with MeOH. The resulting extract was partitioned between EtOAc-MeOH-H₂O (12:1:3). From the EtOAc layer, compound **1a** crystallized. Si gel chromatography of the remaining EtOAc crude fraction allowed the isolation of compounds **2a** and **3a**.

Compound **1a** analyzed for C₂₂H₂₂O₁₁. Its H₂O solubility and a positive Molisch test suggested a glycoside (6). The ir absorption frequencies at 1711, 1613, and 817 cm⁻¹, as well as the uv λ max at 225 and 330 nm, were indicative of a coumarin-like structure (7). The ¹³C-nmr spectra (Table 1) confirmed the presence of 22 carbons, six of which were readily assigned to β -D-galactose (8) and the one at δ 55.9 (q) to a



- 1a** R₁= β -D-Gal, R₂=R₃=H
1b R₁= β -D-Gal, R₂=R₃=Me
1c R₁= β -D-2,3,4,6-tetraacetyl-Gal,
R₂=R₃=Ac
1d R₁=R₂=R₃=H
1e R₁=R₂=R₃=Me
1f R₁=R₂=R₃=Ac
1g R₁=H, R₂=R₃=Me
1h R₁=Ac, R₂=R₃=Me

- 2a** R₁=R₂=R₃=H
2b R₁=R₂=R₃=Me
3a R₁=R₂=Me, R₃=H
3b R₁=R₂=Me, R₃=Ac
4a R₁=Me, R₂=R₃=H

¹For Part II, see Ma T. Reguero, R. Mata, G. Delgado, R. Bye, and E. Linares, *J. Nat. Prod.*, **50**, 315 (1987).

²Dedicated to the memory of the late Dr. Pierre Crabbé.

TABLE 1. ^{13}C -nmr Chemical Shifts of Compounds **1a**, **1d**, **3a** (50 MHz, DMSO- d_6 , TMS as Internal Standard)

Carbon atom	Compounds		
	1a	1d	3a
2	159.52 s	160.00 s	160.90 s
3	112.20 d	110.64 d	100.51 d
4	156.30 s	156.81 s	153.70 s
4a	103.30 s	102.02 s	99.80 s
5a	155.44 s	156.20 s	150.49 s
6	98.50 d	98.30 d	96.41 d
7	162.70 s	162.70 s	163.02 s
8	95.22 d	92.90 d	93.12 d
8a	156.00 s	157.03 s	154.50 s
1'	130.44 s	130.44 s	107.14 s
2'	114.70 d	114.72 d	146.32 s
3'	144.10 s	144.20 s	96.00 d
4'	145.71 s	145.70 s	144.31 s
5'	115.51 d	115.60 d	141.00 s
6'	119.30 d	118.94 d	108.70 d
7-MeO	55.90 q	55.60 q	56.14 q
4'-MeO	—	—	56.14 q
1''	101.12 d	—	—
2''	70.13 d	—	—
3''	73.00 d	—	—
4''	68.10 d	—	—
5''	75.90 d	—	—
6''	60.43 d	—	—

methoxyl group; the remaining ^{13}C signals as well as the singlet at δ 6.20 in the ^1H -nmr spectrum (Table 2) strongly supported that **1a** was the galactoside of a 4-phenylcoumarin (9). Compound **1a** yielded a trimethylether (CH_2N_2), **1b**, and a hexaacetate (Ac_2O /pyridine), **1c**; acid (HCl 1N) and enzymatic hydrolysis (β -galactosidase) afforded β -D-galactose (tlc) and **1d**.

The aglycone **1d** showed important ions at m/z 300 (M^+ , base peak), 272 ($\text{M}^+ - \text{CO}$), and at 257 ($\text{M}^+ - \text{CO} - 15$) (10). The coupling pattern displayed by the aromatic protons (Table 2) and the multiplicities observed for the aromatic carbons in the ^{13}C nmr (Table 1) clearly indicated that one of the benzene rings was trisubstituted, while the other one was tetrasubstituted. The upfield methyl singlets at δ 3.48, 1.59, and 1.42 exhibited in the ^1H -nmr spectra of **1e**, **1f**, and **1h**, respectively, (Table 2) suggested that **1d** had a free hydroxyl group at C-5 (5). Because such a diamagnetic effect was not observed in any of the methyl groups of the -OMe or -OAc functionalities of **1b**, **1c**, and **1g**, it was obvious that the sugar portion was attached to position 5 in **1a**. Thereafter, the placement of the methoxyl group at C-7 in **1a** and **1d** was ascertained by the pyridine and acetylation induced shifts (Table 2) (11). The disposition of the hydroxyl functions at C-4' and C-3' was confirmed by comparison of the spectral and physical properties of **1e** with those previously reported (12), by the chemical shifts observed in the ^{13}C nmr (Table 1), and by the transformation of **1d** to **4a** by treatment with alkali in the presence of air (MeOH/KOH). Compound **4a** was previously isolated from *Hintonia latiflora* (Sesse et Moc. ex DC.) Bullock (13). Finally, the β configuration of the galactosyl unit was inferred by the coupling constant values for the anomeric proton ($J=8$ Hz) as well as for the anomeric carbon [1J $^{13}\text{C}-\text{H}$ (1') = 160 Hz] (14).

Compound **2a** was given the molecular formula $\text{C}_{15}\text{H}_8\text{O}_6$. Its eims showed the

TABLE 2. ¹H-nmr Chemical Shifts of Compounds **1a-1h**, **2a**, **2b**, **3a**, **3b**, and **4a** (80, MHz, TMS as Internal Standard)^a

Compounds	H-3	H-6	H-8	H-2'	H-5'	H-6'	H-3'	OMe				OAc				H-1''	H-2'' H-6''	OH
								C-7	C-5	C-3'	C-4'	C-5'	C-5	C-3'	C-4'			
1a^b	6.20s	7.08d (3)	6.57d (3)	7.38d (3)	7.20d (8)	6.92dd (8,3)	—	3.67s	—	—	—	—	—	—	—	5.26d (8)	4.0- 4.5 m	3.50bs 3.50bs 3.25bs
1b^c	5.95s	6.58s	6.58s	—	6.92bs	—	—	3.85s	3.85s	3.85s	3.90s	—	—	—	—	4.64d (8)	3.0- 3.5 m	—
1c^c	6.15s	6.65d (3)	6.52d (3)	7.10-7.25 m	—	—	—	3.85s	—	—	—	—	—	—	—	5.05d (8)	1.85s, 1.92s 1.99s, 2.18s	—
1c^b	6.05s	6.84d (3)	6.68d (3)	7.20d (3)	7.50d (8)	7.35dd (8,3)	—	3.76s	—	—	2.28s	2.28s	—	—	—	5.16d (8)	3.8- 4.8 m	—
1d^b	6.06s	6.58d (3)	6.48d (3)	7.24d (3)	7.06d (8)	6.90dd (8,3)	—	3.70s	—	—	—	—	—	—	—	5.16d (8)	4.3- 5.7 m	—
1d^d	5.88s	6.25d (3)	6.40d (3)	6.86d (3)	6.87d (8)	6.72dd (8,3)	—	3.85s	—	—	—	—	—	—	—	—	—	9.30bs
1e^c	6.00s	6.23d (3)	6.51d (3)	6.74-6.91 m	—	—	—	3.86s	3.48s	3.92s	3.86s	—	—	—	—	—	—	3.15bs
1f^c	6.07s	6.82d (3)	6.48d (3)	7.10-7.30 m	—	—	—	3.83s	—	—	—	—	—	—	—	—	—	—
1g^c	5.96s	6.27d (3)	6.56d (3)	6.87-7.03 m	—	—	—	3.84s	3.90s	3.90s	—	—	—	—	—	—	—	—
1h^c	6.00s	6.87m (3)	6.46d (3)	6.51-6.93 m	—	—	—	3.87s	3.88s	3.93s	—	—	—	—	—	—	—	—
2a^d	5.91s	6.44s	6.44s	—	7.21s	6.73s	—	—	—	—	—	—	—	—	—	—	—	10.30bs
2b^d	5.90s	6.62s	6.62s	—	7.35s	6.90s	—	3.85s	—	—	3.90s	3.86s	—	—	—	—	—	—
3a^b	5.91s	6.60s	6.60s	—	7.20s	6.90s	—	3.86s	—	—	3.90s	3.86s	—	—	—	—	—	—
3a^b	6.31s	6.55s	6.55s	—	7.56s	6.77s	—	3.70s	—	—	3.78s	3.78s	—	—	—	—	—	—
3b^b	6.31s	6.60s	6.60s	—	7.75s	6.86s	—	3.74s	—	—	3.80s	3.80s	—	—	—	—	—	—
4a^d	5.97s	6.60s	6.60s	—	7.22s	6.73s	—	3.86s	—	—	—	—	—	—	—	—	—	10.35bs 9.15bs

^aCoupling Constants (Hz) in parentheses.^bSolvent pyridine-*d*₅.^cSolvent CDCl₃.^dSolvent CDCl₃/DMSO-*d*₆.

molecular ion at m/z 284 (base peak) and fragment ions at m/z 256 ($M^+ - CO$) and 228 ($M^+ - C_2O_2$). Successive losses of 28 mass units from the molecular ion as well as the singlet at δ 5.91 in the 1H -nmr spectrum (Table 2) and the ir and uv information suggested a 4-phenyl-5,2'-oxido-coumarin skeleton (13). The trimethylether, [2b], obtained by methylation with CH_2N_2 demonstrated the presence of three phenolic hydroxyl groups; the bathochromic shift observed in the uv spectrum upon addition of $AlCl_3$, reversible by HCl, indicated that two of these hydroxyls were in an *ortho* relationship (8). The two one proton singlets at δ 6.73 (H-3') and δ 7.21 (H-6'), as well as the two protons broad singlet at δ 6.44 (H-6, H-8), were consistent with the locations of the hydroxyls at C-4', C-5', and C-7, respectively. Furthermore, the difference in chemical shift between the aromatic protons of 2b and those of 2a ($\Delta H_{6-H_8} = 0.18$, $\Delta H_{3'} = 0.17$, and $\Delta H_{6'} = 0.14$) and the fact that 2b was identical to the methyl derivative of 4a were consonant with the substitution pattern proposed.

The spectral properties of 3a were similar to those of 2a except that the former exhibited a molecular ion at m/z 312 (28 mass units more than 2a) and showed resonances for two methoxyl groups (singlets at δ 3.70 and 3.78) in the 1H -nmr spectrum in pyridine- d_5 (Table 2). Treatment with CH_2N_2 afforded 2b; therefore, the substitution pattern was the same as those in 2a and 4a. The placement of the hydroxyl group in C-5' was finally confirmed by the induced shift, upon acetylation, of one of the aromatic signals of 3a from δ 7.56 (H-6') to δ 7.75 in 3b.

Compound 3a was not toxic to brine shrimp (LC_{50} 1000 ppm, 99% by the IOCD confidence) (15); the biological activity of 1a and 2a is currently under investigation. Compound 1a represents the first 4-arylcoumarin glycoside whose structure has been fully elucidated.

The coexistence of 2a, 3a, and 1a is suggestive of their biogenetic interrelationship, as previously speculated by Bhanu *et al.* (16) during the course of their investigations of chemical transformation of 4-arylcoumarins into xanthenes (16). The biosynthesis of 2a or 3a from 1a might involve an oxidative phenol coupling after hydrolysis with a suitable enzyme.

Regarding the transformation of 1d to 4a it is important to point out that Bhanu *et al.* (16) achieved this kind of transformation by treatment of 5,2' oxygenated 4-phenylcoumarins with boiling HI, and, in that case, the oxide ring formation involved the loss of H_2O between the hydroxyl groups located at positions 2' and 5. However, under basic conditions and in the presence of air, it is not necessary to have any oxygenated substituent at 2' to achieve cyclization. The reaction, thus, might proceed via an oxidative phenol coupling.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—Melting points are uncorrected; ir spectra were recorded on a Nicolet FT-IR instrument, uv spectra were recorded on a Hitachi 220 S double beam spectrophotometer with MeOH as solvent and 1H -nmr spectra on a Varian FT spectrometer in $CDCl_3$, $CDCl_3/DMSO-d_6$, or pyridine- d_5 solutions, with TMS as internal standard; ^{13}C -nmr spectra were taken on a 50 MHz chemagnetics A-200 instrument; mass spectra were determined on a Hewlett-Packard 5985-B spectrometer. Si gel 60 (70-230 mesh) was used for column chromatography; tlc was done on Si gel 60 GF 254 plates (Merck), and the spots were visualized with uv radiation, 0.4 N H_2SO_4 , or anisaldehyde reagent.

PLANT MATERIAL.—The stem bark of *E. caribaeum* was collected in Apaxtla, Guerrero, México, in May 1986. The botanical identification of this material was made by Dr. David Lorence, Instituto de Biología, UNAM. A voucher specimen was deposited at the National Herbarium, (Voucher 25V 1986 col: F. Calzada y E. Castro).

EXTRACTION AND PRELIMINARY FRACTIONATION.—The air-dried, shredded stem bark (4.5 kg) was defatted with hexane. The dried marc was then macerated 3 times with MeOH at room temperature for 3-day periods. The combined MeOH extracts were evaporated to yield a brown residue (1.3657 kg). A portion (382 g) of the MeOH extract was partitioned between EtOAc-MeOH- H_2O (12:1:3), using a continu-

ous liquid-liquid extractor. Evaporation of the organic solvent yielded a residue (104.5 g) that proved, as shown by tlc, to be a complex mixture of products.

ISOLATION OF 5-O- β -D-GALACTOSYL-7-METHOXY-3',4'-DIHYDROXY-4-PHENYLCOUMARIN [1a].—From the organic layer of the partition process, a solid precipitated spontaneously, and this solid, upon repeated recrystallizations from Me₂CO/EtOH, afforded 23.21 g of **1a** (1.84% yield), mp 228–231°; uv λ max (MeOH) 208, 255, 330 nm; ir (KBr) 3650, 3403, 1719, 1613, 1160, 1078, 1049 cm⁻¹. Anal. calcd for C₂₂H₂₂O₁₁: C, 57.14; H, 4.76. Found: C, 56.98; H, 4.89.

ISOLATION OF 7,4',5'-TRIHYDROXY-4-PHENYL-5,2'-OXIDO-COUMARIN [2a] AND 7,4'-DIMETHOXY-5'-HYDROXY-4-PHENYL-5,2'-OXIDO-COUMARIN [3a].—The concentrated organic residue (104.5 g), resulting from the partition process, was chromatographed in a glass column packed with Si gel (1 kg). The initial eluting solvent was CHCl₃ with the percentage of EtOH slowly allowed to increase with time. Fractions of 500 ml each were collected. From fractions 59–73, eluted with CHCl₃-EtOH (99:1), was obtained a yellow, crystalline material, which upon recrystallization with EtOH yielded 542 mg (0.14% yield) of **3a**, mp 273–274°; uv λ max (MeOH) 262, 308, 372, 388 nm; ir (KBr) 3330, 3400, 3175, 3160, 1677, 1618, 1523, 1458, 1289, 1205, 1169, 1120, 817 cm⁻¹; eims *m/z* (rel. int.) 312 (100), 284 (30.1), 269 (10), 256 (51.1). From fractions 328–346, eluted with CHCl₃-EtOH (90:10) was obtained 50 mg (0.013% yield) of **2a**, mp 350° (decomp.); uv λ max (MeOH) 262, 310, 350, 372, 390 nm; (+ AlCl₃) 325, 404 nm; (+ AlCl₃/HCl) 262, 310, 350, 372, 390 nm; ir ν max (KBr) 3425, 3250, 1690, 1630, 1560, 1460, 1298, 1170, 830 cm⁻¹; eims *m/z* (rel. int.) 284 (100), 256 (35), 241 (10), 228 (50).

HYDROLYSIS OF 1a.—Compound **1a** (1 g) was refluxed for 1 h with 100 ml of 1 N HCl. A yellow powder precipitated from the acid solution, and, after washing repeatedly with H₂O and recrystallization from Me₂CO, 600 mg of **1d** was obtained; galactose was identified in the acid solution by comparison with an authentic sample by tlc. Compound **1d**, mp 138–140°; uv λ max (MeOH) 260, 330 nm; ir ν max (KBr) 3440, 1665, 1626, 1598, 1434, 1379, 1294, 1202, 1159, 1082 cm⁻¹; eims *m/z* (rel. int.) 300 (100), 272 (98), 257 (23), 127 (5).

PREPARATION OF METHYLETHER DERIVATIVES 1b, 1e, AND 2b.—Compounds **1a**, **1d**, and **2a** (100 mg each) were dissolved in MeOH; to each solution was added an excess of CH₃N₂ in Et₂O, yielding derivatives **1b**, **1e**, and **2b**, respectively. Compound **1b**, 107 mg, mp 178–180°; ir ν max (KBr) 3400, 1710, 1620, 1430, 1360, 1270, 1258, 1212, 1180, 1080, 860 cm⁻¹. Compound **1e**, 90 mg, mp 166–168°, lit. mp 169–170° (12); uv λ max (MeOH) 252, 328 nm; ir ν max (KBr) 1730, 1620, 1520, 1470, 1420, 1360, 1110, 1020, 820 cm⁻¹; eims *m/z* (rel. int.) 342 (100), 314 (56.9), 299 (10). Compound **2b**, 80 mg, mp 265°, ir ν max (KBr) 1718, 1640, 1622, 1562, 1476, 1430, 1218, 1008, 845 cm⁻¹; eims *m/z* (rel. int.) 326 (100), 298 (45), 270 (60).

ACETYLATION OF 1a, 1d, 1g, AND 3a.—To separate solutions of **1a**, **1d**, **1g**, or **3a** (100 mg each) in 1 ml of pyridine was added 1 ml of Ac₂O; the mixtures were kept at room temperature for 24 h, and after usual work up, the acetyl derivatives **1c**, **1f**, **1h**, and **3b**, respectively, were obtained. Compound **1c**, 100 mg, mp 80–81°; ir ν max (KBr) 1753, 1616, 1506, 1433, 1370, 1219, 1168, 1079, 1055, 903 cm⁻¹. Compound **1f**, 90 mg, mp 169–170°; ir ν max (KBr) 1777, 1763, 1724, 1620, 1506, 1430, 1371, 1345, 1212, 1155, 1068, 908 cm⁻¹. Compound **1g**, oily, 90 mg; ir ν max (KBr) 1780, 1767, 1720, 1640, 1530, 1390, 1210, 1080, 900 cm⁻¹; eims *m/z* (rel. int.) 370 (42), 328 (100), 300 (58), 285 (15), 240 (5), 43 (32). Compound **3b**, 50 mg, oily; ir ν max (KBr) 1766, 1707, 1624, 1506, 1465, 1450, 1279, 1171, 1113 cm⁻¹.

CONVERSION OF 1d TO 4a.—Compound **1d** (25 mg) was treated with 5% KOH-MeOH (10 ml). The reaction mixture was left at room temperature for 3 h. The basic solution was neutralized with 1 N HCl, extracted with two 50-ml portions of EtOAc, and the resulting organic phase was washed twice with H₂O. The extract was concentrated affording an orange residue which upon recrystallization from EtOH yielded 14.7 mg of **4a**, mp 342° (decomp.), lit. mp 342–345° (13); ir ν max (KBr) 3450, 1700, 1640, 1525, 1480, 1313, 1250, 1180 cm⁻¹; uv λ max (MeOH) 260, 310, 375, 390 nm; (+ AlCl₃) 275, 325, 406 nm; (+ AlCl₃/HCl) 260, 310, 375, 390. Compound **1d** (50 mg) was dissolved in a solution of NaOMe (prepared by dissolving 15 mg of Na in 3 ml of anhydrous MeOH). The mixture was stirred at room temperature for 1 h. Evaporation of the solvent gave a residue which was recrystallized from EtOH to yield 15 mg of **4a** (ir and tlc).

CONVERSION OF 1b TO 1g.—Compound **1b** (300 mg) hydrolyzed in the same manner as previously described for **1a**. After work-up, 120 mg of **1g** was obtained; mp 185–187° (EtOH); ir ν max (KBr) 3400, 1700, 1620, 1360, 1160, 1100 cm⁻¹; eims *m/z* (rel. int.) 328 (100), 300 (43), 285 (15), 229 (10).

ENZYMATIC HYDROLYSIS OF **1a**.—Compound **1a** (5 mg) dissolved in H₂O was mixed with β -galactosidase (adequate amounts) and phosphate buffer (pH 4). The mixture was incubated for 12 h at 28°. The completeness of the hydrolysis was monitored by tlc. Galactose was readily identified by tlc in the hydrolysate.

ACKNOWLEDGMENTS

The authors would like to thank the following people: Mr. Alejandro Correa from Negromex, S.A., for the recording of uv and ir spectra; Dr. David Lorence from the Instituto de Biología, UNAM, for the identification of the voucher specimens; the staff of Mass Spectrometry and Magnetic Resonance Laboratories of the Instituto de Química, UNAM, for the recording of several spectra. Also, the authors express their deepest gratitude to Dr. Jerry L. McLaughlin and Ms. Wen-Wen Ma, Purdue University, for the ¹³C-nmr spectra and for the brine shrimp assay of compound **3a**. Finally, sincere thanks are due to Dr. Guillermo Delgado for his continuous support and interest. Special acknowledgment is due to the IOCD for partial support.

LITERATURE CITED

1. W. Standley, "Flora de Guatemala," Field Museum of Natural History, New York, 1975, p. 69.
2. J.T. Roig, "Plantas Medicinales Aromaticas o Venenosas de Cuba," Instituto del Libro, La Habana, 1974, p. 273.
3. F. Villaseñor, *Anal. Inst. Medic. Nac.*, **6**, 194 (1903).
4. K.G. Krebs and N.E. Griesinger, *Arzneimitt. Forsch.*, **10**, 32 (1960).
5. F. Sanchez-Viesca, *Phytochemistry*, **8**, 1821 (1969).
6. H.T. Clarke, "Organic Analysis," Arnold, London, 1966, p. 92.
7. D.M.X. Donnelly, J.C. Thompson, W.B. Whalley, and S. Ahmad, *J. Chem. Soc., Perkin I*, 1737 (1973).
8. K.R. Markham, "Techniques of Flavonoid Identification," Academic Press, New York, 1982, pp. 45, 81.
9. A. Pelter, R.S. Ward, and T. Gray, *J. Chem. Soc., Perkin I*, 2475 (1976).
10. B.J. Donnelly, D.M.X. Donnelly, and A.M. O'Sullivan, *Tetrahedron*, **24**, 2617 (1968).
11. R. Alves de Lima, G. Delle Monache, and B. Botta, *Rev. Latinoamer. Quim.*, **13**, 61 (1982).
12. G. Delle Monache, B. Botta, A.S. Neto, and R. Alves de Lima, *Phytochemistry*, **22**, 1657 (1983).
13. G. Reher and L. Kraus, *J. Nat. Prod.*, **47**, 172 (1984).
14. K. Bock and C. Pedersen, *J. Chem. Soc., Perkin Trans. II*, 293 (1974).
15. B.N. Meyer, N.R. Ferrigni, J.E. Putnam, L.B. Jacobsen, D.E. Nichols, and J.L. McLaughlin, *Planta Med.*, **45**, 31 (1982).
16. S. Bhanu, T. Saroja, T.R. Seshadri, and S.K. Mukerjee, *Indian J. Chem.*, **10**, 577 (1972).

Received 16 February 1987